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## 5'-Homoaristeromycin. Synthesis and antiviral activity against orthopox viruses

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**Abstract**—An efficient synthesis of 5'-homoaristeromycin has been developed. This permitted an extensive antiviral analysis, which found potent activity toward vaccinia, cowpox, and monkeypox viruses. For comparative purposes, 5'-homoadenosine was made available by a newly designed route and found to be inactive.

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Inhibitors of S-adenosyl-L-homocysteine (AdoHcy) hydrolase have shown promise as antiviral agents<sup>1,2</sup> by disrupting essential viral macromolecular methylation processes.<sup>2</sup> Carbocyclic nucleosides <sup>3</sup> represent a prominent class of compounds whose antiviral potential has been traced to such an effect.<sup>1</sup> Within that group, carbocyclic adenosine (aristeromycin, 1) is at the center of these investigations<sup>4</sup> but its promise is limited by a toxicity arising from 5'-phosphate formation.<sup>5</sup>

Structural modifications of **1** with the aim of reducing phosphate-based toxicity have yielded meaningful drug candidates.<sup>6</sup> An approach not explored, however, is extension of the C-5' hydroxymethyl side chain by a methylene group to provide the C-5' homolog of aristeromycin (**2**). This analog can be expected<sup>7</sup> to have displaced the phosphate-susceptible hydroxyl from the phosphate-transfer zone in the kinases responsible for metabolism to **1** to its nucleotides. In support of this, **2** has been reported<sup>8</sup> to be inactive against HSV-1 and HSV-2, possibly, due to its failure to be phosphorylated (Fig. 1).

To investigate 2 more thoroughly as a possible antiviral agent a more practical synthesis of it was necessary. For comparative antiviral purposes, 5'-homoadenosine (3) was also sought by a much more efficient way than exists

3, X=O, n=2

Figure 1.

in the literature. The results of this effort are communicated here.

Existing methods<sup>8,10,11</sup> for preparing 5'-homoaristeromycin (2) suffer from too many steps, limited scale-up, low yields and, in one case, resulting in a racemic product. Therefore, an efficient and stereoselective synthesis of 2 was needed. Starting from enone 4<sup>12</sup> (Scheme 1), 1,4-addition of ethyl trimethylsilylacetate followed by in situ cleavage of the trimethylsilyl group furnished, stereoselectively, the ketone ester 5 as the only product. The stereochemistry at C-4 of 5 was derived from the fact (i) that a 1,4-addition to the concave structure of **5** can be expected  $^{13}$  to give a  $\beta$  (up) product and (ii) that 5 was converted into the known 2. Reduction of 5 with sodium borohydride provided the coupling precursor 6, which has been reported via a more tedious way.14 Mitsunubo coupling reaction of 6 with 6-chloropurine furnished 7. Selective reduction of 7 with diisobutylaluminum hydride (DIBAL) yielded the desired alcohol 8.

Keywords: Carbocyclic nucleosides; Nucleoside homologation; Monkeypox.

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EtO<sub>2</sub>C

$$A^{12}$$
 $C$ 

EtO<sub>2</sub>C

 $A^{12}$ 
 $C$ 
 $A^{12}$ 
 $C$ 
 $A^{12}$ 
 $C$ 
 $A^{12}$ 
 $A^{12}$ 

Scheme 1. Reagents and conditions: (a) (i) ethyl (trimethylsilyl)acetate, *n*-BuLi, DIPA, HMPA/THF, -78 °C, (ii) KF, EtOH/H<sub>2</sub>O, rt, 84% for two steps from 4; (b) NaBH<sub>4</sub>, MeOH, ice temp, 100%; (c) 6-chloropurine, Ph<sub>3</sub>P, DIAD, THF, 0–50 °C, 52%; (d) DIBAL, CH<sub>2</sub>Cl<sub>2</sub>, -50 °C, 80%; (e) NH<sub>3</sub>/MeOH, 94%; (f) HCl/MeOH, 95%.

Scheme 2. Reagents and conditions: (a) (i) DMSO, DIPEA, SO<sub>3</sub>-Py, CH<sub>2</sub>Cl<sub>2</sub>, 90%, (ii) Ph<sub>3</sub>PCH<sub>3</sub>Br, *t*-BuOK, Et<sub>2</sub>O, 75%; (b) 9-BBN, THF, then NaOH, H<sub>2</sub>O<sub>2</sub>, 97%; (c) (i) 70% AcOH, 85°C; (ii) Ac<sub>2</sub>O, DMAP, pyridine, 85% for two steps from 12; (d) Adpiv, HMDS, TMSCl, TMSOTf, 60%; (e) NH<sub>3</sub>/MeOH, 91%.

Ammonolysis of **8** (to **9**) followed by hydrolytic deprotection smoothly afforded **2** in good overall yield. <sup>15</sup>

The known syntheses of 6'-homoadenosine<sup>9</sup> (3) either involved many steps<sup>9a</sup> or suffer a low yield of the final product.<sup>9b,c</sup> Our plan (Scheme 2) envisioned beginning with homologation of 10.<sup>16</sup> Side chain oxidation of 10 followed by Wittig olefination afforded 11. Submitting 11 to regioselective hydroboration with 9-BBN followed by oxidative hydrolysis, smoothly provided 12 in high yield. Hydrolysis of 12 with acetic acid produced a tetrol, which was fully protected with acetic anhydride to provide the anomeric acetate 13. The coupling reaction of 13 with *N*-pivaloyl protected adenine under Vorbrüggen glycosylation conditions yielded the desired N-9 product 14 as the only isolated product. Deprotection of 14 with ammonia furnished 3 in good overall yield<sup>17</sup> from D-ribose.

Compounds **2** and **3** were evaluated against a wide variety of both DNA viruses and RNA viruses. <sup>18</sup> From this, very significant effects were seen for **2** toward vaccinia (IC<sub>50</sub> 1.2  $\mu$ g/mL), cowpox (IC<sub>50</sub> 0.12  $\mu$ g/mL), and monkeypox (IC<sub>50</sub> 0.12  $\mu$ g/mL) viruses, all in Vero 76 cells with CC<sub>50</sub> > 100  $\mu$ g/mL. This observation is particularly noteworthy since it is well known that vaccinia is susceptible to AdoHcy hydrolase inhibitors but cowpox was thought not to be. <sup>18e</sup> In any case, details of this investigation and the, possibly, less notable activity of **2** toward other viruses <sup>18</sup> will be forthcoming, including its potency toward variola. <sup>19</sup> Analog **3** was inactive in all of the assays employed. <sup>18</sup>

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- 15. Selected data for **2**: mp 178–179 °C; <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  8.20 (s, 1H), 8.11 (s, 1H), 7.17 (s, 2H), 4.90 (d, J = 6.27 Hz, 1H), 4.68 (d, J = 4.63 Hz, 1H), 4.57 (m, 1H), 4.43 (t, J = 5.12 Hz, 1H), 4.35 (m, 1H), 3.71 (m, 1H), 3.46 (m, 2H), 2.24 (m, 1H), 1.97 (m, 1H), 1.74 (m, 2H), 1.58 (m, 1H); <sup>8</sup> <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  156.0, 152.1, 149.6, 140.2, 119.3, 74.9, 74.3, 59.9, 59.4, 40.0, 37.4, 32.5. Anal. Calcd for C<sub>12</sub>H<sub>17</sub>N<sub>5</sub>O<sub>3</sub>: C, 51.60; H, 6.14; N, 25.08. Found: C, 51.39; H, 6.18; N, 24.81.
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- 17. Selected data for 3: mp 223–224 °C (lit. 9c 231.5–232.5 °C); 

  <sup>1</sup>H NMR (250 MHz, DMSO- $d_6$ )  $\delta$  8.27 (s, 1H), 8.10 (s, 1H), 7.24 (s, 2H), 5.79 (d, J = 5.3 Hz, 1H), 5.36 (d, J = 5.7 Hz, 1H), 5.11 (d, J = 5.1 Hz, 1H), 4.61 (m, 1H), 4.44 (t, J = 5.1 Hz, 1H), 4.03 (m, 1H), 3.93 (m, 1H), 3.42 (m, 2H), 1.76 (m, 2H); <sup>13</sup>C NMR (62.9 MHz, DMSO- $d_6$ )  $\delta$  156.1, 152.6, 149.4, 139.9, 119.2, 87.4, 81.0, 73.4, 72.9, 57.5, 36.6. Anal. Calcd for C<sub>11</sub>H<sub>15</sub>N<sub>5</sub>O<sub>4</sub>: C, 46.97; H, 5.38; N, 24.90. Found: C, 46.85; H, 5.48; N, 24.63.
- For leading references on the procedures used for the assays see (a) Ref. 5; (b) Siddiqi, S. M.; Chen, X.; Schneller, S. W.; Ikeda, S.; Snoeck, R.; Andrei, G.; Balzarini, J.; De Clercq, E. J. Med. Chem. 1994, 37, 551; (c) Seley, K. L.; Schneller, S. W.; Korba, B. Nucleosides Nucleotides 1997, 16, 2095; (d) http://www.usu.edu/iar/Brochure/brochure.html (September 6, 2004); and; (e) Baker, R. O.; Bray, M.; Huggins, J. W. Antiviral Res. 2003, 57, 13.
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